

Expert Opinion

1. Introduction
2. Body
3. Conclusion
4. Expert opinion

What should be considered on design of a colon-specific prodrug?

Yunjin Jung[†] & Young Mi Kim

Pusan National University, College of Pharmacy, Laboratory of Biomedicinal/Medicinal Chemistry, Busan, 609-735 Korea

Importance of the field: Generally, a prodrug, a pharmacologically inactive derivative of an active drug, is designed to modulate pharmacokinetic properties of the parent drug. Targeted distribution of an orally administered drug at the large intestine confers therapeutic advantages on treatment of colonic diseases, peptide and protein therapy and chronotherapy.

Areas covered in this review: To achieve such distribution control in the gastrointestinal tract, the adoption of the prodrug concept gives birth to a colon-specific prodrug. The requirement for a prodrug to be colon-specific is described along with the necessary and sufficient conditions of drugs for conversion to a colon-specific prodrug. The known and previously unnoticed factors that negatively influence therapeutic activity and reproducibility of a colon-specific prodrug are presented with suggestions to minimize the negative influence.

What the reader will gain: This review provides tactics to satisfy the requirements for being colon-specific and the potential strategies to circumvent obstacles in developing an efficient colon-specific prodrug.

Take home message: On design of a colon-specific prodrug, one should take into consideration not only delivery of a drug to the target site, but also the therapeutic effectiveness there.

Keywords: colon-specific carrier, colon-specific prodrug, colonic metabolism, drug delivery, intestinal microflora, oral drug delivery, prodrug and site-specific drug delivery

Expert Opin. Drug Deliv. (2010) 7(2):245-258

1. Introduction

Site-specific delivery of a drug is a useful strategy to improve therapeutic properties of the drug [1,2]. Unlike the conventional way of drugs traveling through the body, which usually leads to extensive distribution, site-specific delivery is aimed at a site where there is a therapeutic target [2]. This site-restricting control of drug distribution enables the therapeutic concentration of a drug to be reached at a target site while its blood concentration is kept low, which should result in an increase of the therapeutic potency as well as a decrease of side effects. For such advantages, delivery systems targeting a variety of tissues have been developed [3-11].

Colon-specific drug delivery implies transport of a drug to the large intestine without significant loss in the upper intestine on oral administration of the drug [12]. Colonic delivery is directed mainly at achieving efficient treatment of diseases that develop at the colonic site and reducing side effects by restricting systemic absorption [13,14]. Conventional dosage forms are designed to be disintegrated, dissolved and subsequently absorbed systemically in the upper intestine. The colon, where activity of proteolytic enzymes is lower than in the small intestine, could also be utilized as the site of absorption for those drugs that are unstable in the upper intestine, such as therapeutic peptides and proteins [15,16]. Colonic targeting may be

informa
healthcare

Article highlights.

- Restriction of the systemic absorption of a drug in the upper intestine is linked to an efficient delivery of the drug to the large intestine.
- A great metabolic activity of microflora in the colon could be exploited for colon-specific activation of a prodrug.
- The metabolic activity of microflora in the colon could be variable among individuals and even in a single subject.
- All therapeutic agents that are applicable to treatment of colonic diseases, peptide and protein therapy and chronotherapy could be candidates for modification to colon-specific prodrugs.
- For a drug to become a colon-specific prodrug, an appropriate functional group is a prerequisite for conjugation with a colon-specific promoiety.
- Colonic distribution of the parent drug after oral administration of a colon-specific prodrug is relevant to an efficient therapeutic action against a colonic disease whose pathogenic lesion occurs not only in the proximal part but also in the distal part of the large intestine.
- The rate of bacterial enzyme-mediated conversion of colon-specific prodrugs to the parent drugs is one of the determinants to control regional therapeutic availability of the drugs in the large intestine.
- Colonic metabolic stability of drugs is one of the determinants to control regional therapeutic availability of the drugs in the large intestine.
- Insightful suggestions and potential strategies for design of an efficient colon-specific prodrug are provided.

This box summarises key points contained in the article.

applicable to delaying absorption to achieve a desirable time–concentration profile for the treatment of diseases such as asthma, gastric ulcer and arthritis, which often have peak symptoms at bedtime [17–19].

Two major approaches available for colon-specific drug delivery are pharmaceutical formulation and prodrug [20]. The pharmaceutical formulations for colonic delivery include coating or tableting with pH-sensitive or colonic (microbial) enzyme-degradable polymers and the design of timed-release dosage forms [21]. The prodrug approach is achieved by coupling a hydrophilic (or polymeric) and enzymatically disposable carrier to a candidate drug, which renders the coupled product (prodrug) non-absorbable in the upper intestine and cleavable to liberate the drug in the large intestine [22]. Generally, although applicable limitedly to drugs with a chemical group available for coupling with a colon-specific carrier, the prodrug approach confers a drug better colon-targetability than does the pharmaceutical approach.

2. Body

2.1 Restriction of systemic absorption

For a foreign compound ingested to reach the large intestine without significant systemic absorption in the upper intestine, the compound should be hydrophilic or large, which prevents passive transport through the intestinal epithelial cell

layer [23,24]. Conjugation of a parent drug with hydrophilic small molecules such as sulfate, amino acids and sugars is a way to restrict systemic absorption of the drug in the upper intestine [25–27]. Another way is to conjugate with various natural or synthetic polymers. Some polymers used for this purpose, such as cyclodextrin, dextran, poly(L-aspartic acid), have biodegradable backbones susceptible to colonic bacterial enzymes [28–30]. Others, such as sulfaniamidoethylene polymer, N-(hydroxypropyl)methacrylamide copolymers and polyamidoamine dendrimer, are not degradable [31–33]. In addition, the linkage between the drug and the promoiety should be chemically and enzymatically stable during the transit through the stomach and small intestine.

2.2 Colon-specific activation of a prodrug

The intestinal microflora has a great metabolic activity participating in the metabolism of exogenous and endogenous compounds, which has a metabolic potential equal to or greater than that of the liver [34]. The number of bacteria in stomach and proximal small intestine ($10^2 - 10^4$ cfu/ml) is similar and increases to $10^5 - 10^7$ cfu/ml in the lower part of the small intestine. In the colon, the bacterial count reaches $10^{11} - 10^{12}$ cfu/ml, where the anaerobic bacteria outnumber aerobes by a factor of $10^2 - 10^4$ [35]. The ‘atmospheric’ condition in the colon is a total lack of oxygen, rendering the oxidation–reduction potential very low, which is in the range -200 to -250 mV [35,36]. As summarized in Table 1, differences between metabolisms in gut flora and liver are the accessibility and the type of predominant metabolic reactions [37]. In general, oxidation and conjugation are the predominant metabolic reactions in the liver, whereas reduction and hydrolysis are the preferred ones in the large intestinal lumen. This unique environment of the colon could be exploited for colon-specific activation of a prodrug, which is a pharmacologically inactive version of a parent drug. Typical drug metabolizing enzymes in human intestinal microflora, some of which have been utilized for colon-specific activation of prodrugs, are listed in Table 2 [37]. Several reviews deal with colonic drug delivery using the prodrug strategy, in which prodrugs are introduced as being classified by promoieties or bacterial enzymes [20,22,38].

2.3 Variation in microflora

Microbially triggered activation of colon-specific prodrugs seems to be a solid strategy for the release of parent drugs from the prodrugs at the target site. However, there are some issues that should be considered for the therapeutic feasibility of the prodrugs. Inter-individual difference in microflora is one of the issues. Considerable variations in the composition of the microflora are found among individuals [35], which might result in variation in therapeutic activity of a colon-specific prodrug. In a single healthy subject, intestinal flora is quite stable over a prolonged period of time [35]; but, even in a single subject, disease states are reported to influence intestinal

Table 1. Differences between metabolism in colonic microflora and liver.

	Colonic flora	Liver
Accessibility	Limited*	All drugs systemically available
<i>Phase I metabolism</i>		
Oxidation	Absent	Predominant
Reduction	Predominant	Unimportant
Hydrolysis	Important (more)	Important
<i>Phase II metabolism</i>		
	Cleavage of Conjugation	Conjugation

*Cases for a drug to reach colonic microflora: poor intestinal absorption, biliary excretion and colon-specific delivery.

Table 2. Drug-metabolizing enzymes in the human colonic microflora.

Enzyme	Metabolic reaction catalyzed
<i>Hydrolytic reaction</i>	
Esterase and amidase	Cleavage of esters or amides of carboxylic acids
Glycosidase	Cleavage of β -glycosides of alcohols and phenols
Glucuronidase	Cleavage of β -glycosides of alcohols and phenols
Sulfatase	Cleavage of O-sulfates and sulfamates
<i>Reductive reaction</i>	
Nitroreductase	Reduction of aromatic and heterocyclic nitro compounds
Azoreductase	Reductive cleavage of azo compounds
N-oxide reductase	Removal of oxygen from N-oxide
Sulfoxide reductase	Removal of oxygen from sulfoxides
Hydrogenase	Reduction of carbonyl groups and aliphatic double bonds

flora [35,39]. Crohn's disease patients show increased *Escherichia coli* and lactobacilli in the large intestine whereas ulcerative colitis patients do not [40]. Indeed, a decrease in glycosidase and azoreductase activity is observed in the feces of patients with active Crohn's disease [41]. The disease states accompanying diarrhea, such as cholera, tropical sprue and irritable bowel disease, tend to increase aerobic bacteria or decrease anaerobic bacteria concentration in the fecal flora [35,42].

2.4 Candidate drugs for modification to colon-specific prodrugs

As described earlier, efficient treatment of colonic diseases including inflammatory bowel disease (IBD), systemic

absorption of therapeutic agents that are not stable in the upper intestine, or modulation of time-concentration to match time-symptoms for chronotherapy are the cases appropriate for colon-specific prodrug approaches [18]. All drugs that are medicated to colonic diseases, IBD, constipation/diarrhea, colorectal cancer, irritable bowel disease and intestinal infection could be candidates for modification to colon-specific prodrugs, which should increase the potency and decrease the systemic side effects. Anti-inflammatory drugs, 5-aminosalicylic acid (5-ASA) and glucocorticoids, which are used commonly for treatment of IBD [43], were subjected to conjugation with hydrophilic small molecules or natural and synthetic polymers to be delivered to the large intestine [22]. In addition, colon-specific prodrugs of metronidazole, nalidixic acid and sulfa drugs (antimicrobial agents), menthol (antispasmodic agent), naloxone and nalmefene (anti-constipation), ursodeoxycholic acid (chemopreventive agent), 9-aminocampothecin and 5-fluorouracil (anticancer agents) were reported [32,44-50]. Naturally occurring glycosides, rutin and sennosides, act as colon-specific prodrugs that deliver to the large intestine and liberate their aglycons, quercetin and rhein anthrone, followed by exerting their respective biological activities, anti-inflammatory and laxative effects [51,52]. Besides the above colon-specific prodrugs for treatment of colonic diseases, immunosuppressive agents (treatment of IBD), vermicides (treatment of intestinal parasites) and COX-2 selective inhibitors (prevention of colorectal cancer) are candidate drugs for modification to colon-specific prodrugs. In fact, colonic delivery of mebendazole, a vermicide, and celecoxib, a selective COX-2 inhibitor, using pharmaceutical formulations has been reported [53-55]. No report has been published on a prodrug for colonic delivery of peptide and protein drugs, while vasopressin, insulin and calcitonin were pharmaceutically formulated to be delivered to the large intestine for systemic absorption [16,56,57]. A capsule or pellet containing peptide therapeutics was coated with synthetic polymers carrying an azo bond in the backbones, which are cleavable by bacterial azoreductase in the colon [58]. Harboe *et al.* investigated pharmacokinetics of various dextran-naproxen ester conjugates administered orally in animals and found that the naproxen plasma profiles for all the polymeric prodrugs showed a characteristic lag time of naproxen appearance in the blood (2–3 h) compared with the administration of the parent drug [29]. Similar delayed absorption of ketoprofen was observed in the study on various dextran-ketoprofen ester conjugates [59]. The authors reasoned that the delay in systemic absorption is due to colonic delivery of the dextran conjugates and subsequent release of the parent drugs from the conjugates. The delayed systemic absorption may be advantageous to the management of symptoms of arthritis, asthma and gastric ulcer, which may commonly have a peak symptom at bedtime. Therefore, therapeutic agents for the treatment of such diseases should also be potential candidate drugs for modification to colon-specific prodrugs. These include non-steroidal anti-inflammatory agents,

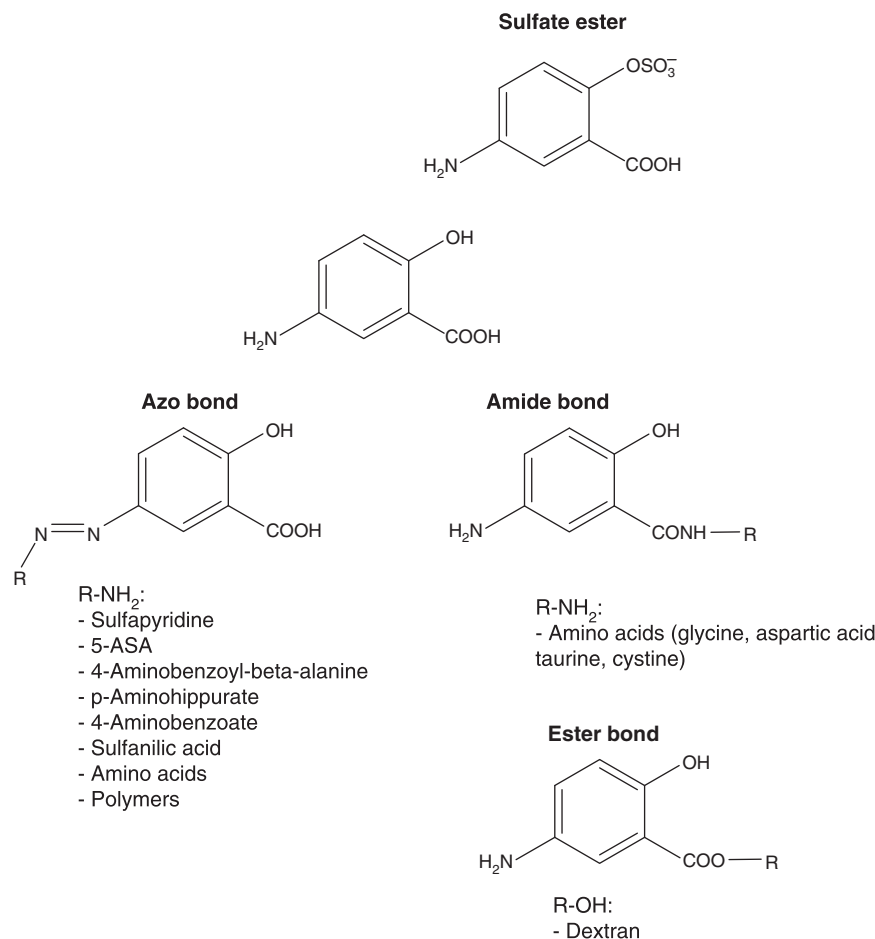


Figure 1. Various chemical modifications imposed on 5-aminosalicylic acid to couple colon-specific promoieties to 5-aminosalicylic acid.

histamine receptor 2 blockers, proton pump inhibitors and antiasthmatic agents.

2.5 Functional groups appropriate for coupling with a colon-specific promoiety

Candidate drugs have been introduced above, which should become more efficient in therapeutic aspects when modified to a colon-specific prodrug. Unfortunately, not all the candidate drugs can be modified to such a prodrug. For a drug to become a colon-specific prodrug, an appropriate functional group is a prerequisite for conjugation with a colon-specific promoiety, where the covalent link should be stable in the upper intestine and cleavable in the large intestine.

5-Aminosalicylic acid, an anti-inflammatory agent, is the mainstay for inducing and maintaining remission in mild-to-moderate IBD such as ulcerative colitis and Crohn's disease [43]. As shown in Figure 1, various chemical modifications have been imposed to 5-ASA to deliver the drug specifically to the target site [22]. Three functional groups in 5-ASA, aromatic amine, carboxylic group and phenol, have been utilized for conjugation with a promoiety. The aromatic amine was

conjugated with sulfapyridine, another 5-ASA, 4-aminobenzoyl-β-alanine, *p*-aminohippurate, 4-aminobenzoate, sulfanilic acid and amino acids via an azo bond, namely, sulfasalazine, olsalazine, balsalazide, ipsalazide, bensalazide, salicylazosulfanilic acid and 5-ASA-amino acid azo conjugates, respectively [60-63]. Polymers with a pendant group that is able to form an azo bond on reaction with 5-ASA were reported as polymeric colon-specific prodrugs of 5-ASA [31,33,64]. Recently, 5-ASA was conjugated with diamino-poly(ethylene glycol) by means of an azo bond for colon-specific delivery [65]. The physicochemical nature of the polymeric 5-ASA prodrugs depends on monomers consisting of the polymers, which affects enzymatic release of 5-ASA [66]. 5-ASA derivatives where the amine was conjugated with the carboxylic group in dipeptides were developed. However, they were designed to release 5-ASA in the small intestine [67]. The carboxylic group in 5-ASA was coupled with amino acids by means of an amide bond, which is susceptible to colonic bacterial amidase but not to host ones in the upper intestine [26]. In the case of using amino acids as a promoiety, some amino acids such as taurine, cysteine and phenylalanine [63,68,69] play dual roles, not only

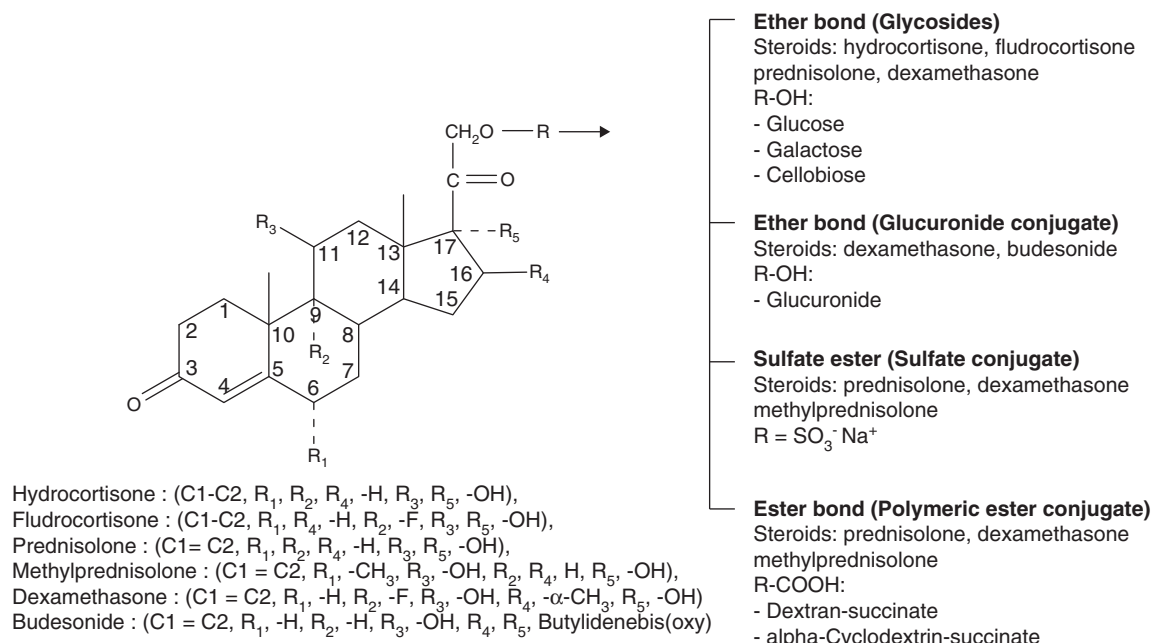


Figure 2. Various chemical modifications imposed on glucocorticoids to couple colon-specific promoieties to the glucocorticoids.

delivering 5-ASA to the large intestine but also enhancing the therapeutic effects of 5-ASA on experimental colitis. Kim *et al.* revealed a molecular mechanism for the taurine effect, which an additive inhibitory effect of 5-ASA and taurine on the inflammatory transcription factor, NF-κB, was involved in [70]. The carboxylic acid was conjugated with dextran by means of an ester bond [71]. Although an ester bond is not a linkage suitable for colon-specific delivery considering that esterases are abundant in the whole gastrointestinal (GI) tract without significant regional preference [72], dextran-drug ester conjugates including dextran-5-ASA ester conjugates show a good colon-targetability. Larsen *et al.* suggest that the ester bond between drugs and the polymeric carrier dextran is protected by steric hindrance from esterase attack in the upper intestine, whereas the ester bond is exposed by degradation of the dextran backbone to oligomers by bacterial dextranases in the colon, resulting in hydrolysis and liberation of parent drugs [73]. The phenol group in 5-ASA was also used for coupling with a promoiety. The phenol was sulfated to afford 5-ASA sulfate ester, which should be more hydrophilic than 5-ASA and is desulfated in the colonic contents while stable in the contents of the upper intestine, thereby satisfying the requirement for colon targeting [74].

Glucocorticoids, which have been used for attack therapy against IBD, are well absorbed in the upper intestine, causing serious side effects on long-term administration [75]. Thus, development of a colon-specific delivery system for a glucocorticoid is highly desirable. The hydroxyl groups in glucocorticoids are the sites for coupling with a promoiety,

as shown in Figure 2. Sugars such as glucose, galactose and cellobiose were attached to the hydroxyl groups of glucocorticoids to afford glycosides [27,76]. The glycosidic ether linkages between sugars and glucocorticoids are cleaved to release the active drugs by bacterial glycosidases in the colon, whose release rate depends on sugars [76]. Luminal glucosidase activity is present in the small intestine, which is from sloughed mucosa cells in the GI tract [77]. However, glycoside prodrugs do not seem to be degraded to a significant extent in the small intestine because the glucosidase activity is lower than that in the colon [76]. Dexamethasone and budesonide-glucuronide conjugates were introduced as colon-specific prodrugs, where the hydroxyl groups of the steroids were modified to glucuronide ether linkages [78,79]. As luminal β-D-glucuronidase activity has greater regional preference to the colon than luminal β-D-glucosidase activity, the glucuronide prodrugs are suggested to have better colonic delivery efficiency than glucoside prodrugs [78]. Doh and co-workers added sulfate ester conjugation to modifications of glucocorticoids for colon-specific prodrugs [25,80]. Sulfated metabolites excreted through the bile duct are known to be desulfated by bacterial sulfatases in the large intestine. This colonic metabolism was taken advantage of to deliver a chemopreventive agent, ursodeoxycholic acid (UDCA), to the colon. Rodrigues *et al.* reported that sulfated UDCA, which is greatly hydrophilic, was delivered specifically to the large intestine without significant loss in the upper intestine compared with UDCA [49]. In the same way, sulfation of glucocorticoids renders the parent drugs more hydrophilic, limiting their systemic

absorption, which is linked to efficient delivery to the large intestine [81]. Side effects including adrenal suppression are attenuated significantly comparing the sulfate prodrugs with the parent steroids, whereas the potency of the prodrugs is greater than that of the corresponding steroids [82,83]. Dextran is also used as a colon-specific promoiety for glucocorticoids. The hydroxyl groups in the drugs and the carrier are conjugated by means of a linker molecule, succinic acid, where esters are formed at both ends of the linker molecule. The glucocorticoid-succinate-dextran conjugates act as a polymeric colon-specific prodrug and show fewer side effects and greater potency than the corresponding steroids [84,85]. Like dextran-succinate-glucocorticoid conjugates, α -cyclodextrin-succinate-prednisolone conjugate, where α -cyclodextrin was used as a carrier for colon-specific delivery of the steroid, possesses similar advantages over the parent drug [86,87].

5-Fluorouracil (5-FU), used for a variety of cancers including colorectal cancer, is administered parenterally because oral administration of the drug shows low and variable bioavailability owing to the incomplete absorption and marked bioinactivation by dihydrouracil dehydrogenase in the liver and the mucosal membrane of the GI tract [88]. Bioactivation of 5-FU to 5-fluoro-2'-deoxyuridine was greater in colon cancer than in normal tissue whereas bioinactivation is minimal in the colon and colon tumor, suggesting that colon-specific delivery of 5-FU should improve the therapeutic property of 5-FU [89,90]. The nitrogens in the pyrimidine ring of 5-FU might be available for secondary amide formation but not for diazo reaction. Thus, amino acid conjugation by means of an amide bond may be a feasible way to design a colon-specific prodrug of 5-FU, which remains to verify whether the secondary amide acts as a colon-specific linkage and the steric hindrance does not prevent enzymatic activation. To avoid such uncertainty, Lee *et al.* designed *N*-Acyl-2-acetoxy-DL-glycine (NAG) as a chemical drug delivery system for colonic delivery of 5-FU (Figure 3A) [50]. The pyrimidine nitrogen (N-1 position) in 5-FU was substituted with the acetoxy group to afford *N*-nicotinoyl-2-(5-fluorouracil-1-yl)-DL-glycine (NFG), in which the nicotinoyl amide is proposed to be cleaved by bacterial enzymes, immediately followed by spontaneous degradation resulting in release of 5-FU in the large intestine, as shown in Figure 3B. It was found that the release per cent of 5-FU in the cecal contents is low (9% of the dose), probably owing to the steric hindrance imposed by 5-FU against enzymatic cleavage of the amide bond. Another glycol moiety was inserted next to the nicotinoyl group of NFG to decrease the steric hindrance, expecting enhanced hydrolysis of nicotinoyl group to release 5-FU more readily than NFG. [91]. The 5-FU prodrug (Figure 3C) liberated 5-FU in the cecal contents, which increased 5-FU release up to 16% of the dose. The colon-specific chemical delivery system, NAG, was applied to colon-specific delivery of an amebicidal agent, metronidazole (MTZ) [44]. The hydroxyl group in MTZ was easily substituted with the acetoxy group to produce a colon-specific prodrug of

MTZ, *N*-nicotinoyl-2-{2-(2-methyl-5-nitroimidazole-1-yl)ethoxy}glycine (Figure 3D), whose release mechanism is thought to be the same as that of the 5-FU prodrug. In *in vitro* and *in vivo* experiments, the MTZ prodrug was shown to be delivered to and released MTZ specifically at the large intestine. The authors suggest that the colon-specific chemical delivery system should be applicable to any drugs with an appropriate nucleophile that is able to be substituted with the acetoxy group.

2.6 Colonic distribution of the parent drugs after oral administration of colon-specific prodrugs

For an efficient therapeutic action against a colonic disease such as inflammatory bowel disease, a colon-specific prodrug is required to deliver an active agent not only to the proximal part but also to the distal part of the large intestine where inflammatory lesion of target diseases frequently occurs [14]. In this respect, the colonic metabolic susceptibility of both a parent drug and its prodrug should be considered on design of a colon-specific prodrug. If the prodrug is very susceptible to colonic metabolism(s), almost all of it would be converted to the parent drug at the proximal part of the large intestine and would not reach the distal large intestine. In this case, it might depend on the colonic metabolic stability of the parent drug whether the drug could reach the distal large intestine. If the prodrug has an appropriate half-life for the metabolic conversion, substantial amount of the prodrug would arrive at and be converted to the active agent in the distal large intestine, thus probably affording the active agent regardless of the metabolic stability of the active agent. Of course, the more stable the active agent, the more it would be accumulated at the distal large intestine. Therefore, it is thought to be important to select an active agent and a colon-specific promoiety that have a proper metabolic property for design of an efficient colon-specific prodrug. The following two sections deal with more details on these issues.

2.6.1 Efficiency of bacterial enzyme-mediated activation of a prodrug

This issue involves the rate of bacterial enzyme-mediated conversion of colon-specific prodrugs to the parent drugs, which would affect regional therapeutic availability of the parent drugs in the large intestine after oral administration of colon-specific prodrugs. The rate of a colon-specific prodrug conversion to a parent drug depends mainly on susceptibility of the drug-promoiety linkages to colonic metabolisms. Even in the case of the same linkage, the susceptibility should vary depending on promoieties and the parent drugs. The conversion rates of 5-ASA-amino acid conjugates were compared varying the promoiety, amino acids. The order of the conversion rates is 5-ASA-glycine > 5-ASA-tyrosine > 5-ASA-aspartic acid in the cecal contents [92], which is in a parallel pattern with fecal recovery of 5-ASA and the prodrugs [68]. The fecal recovery of the prodrugs increases as the *in vitro*

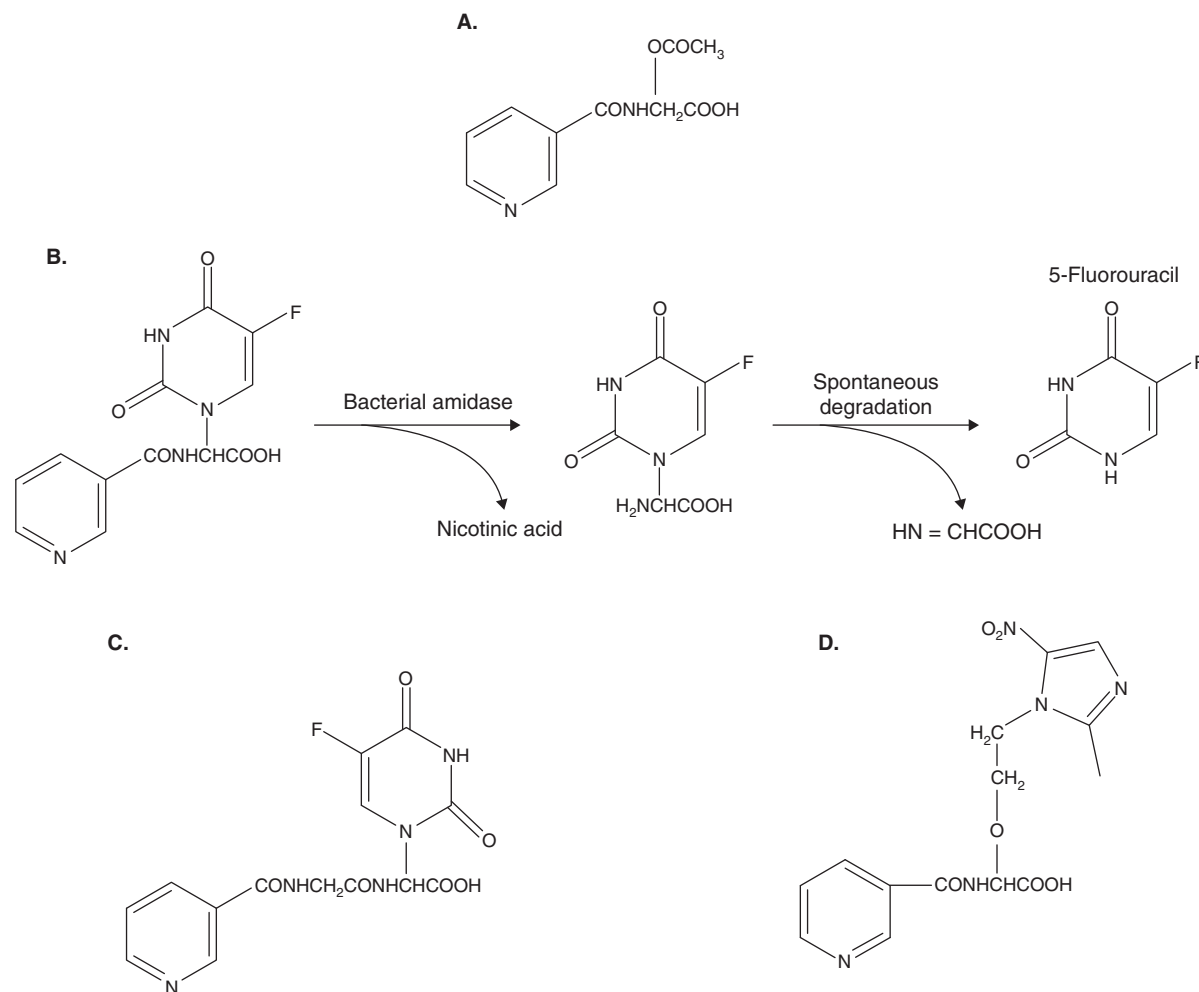


Figure 3. A colon specific chemical drug delivery system of 5-fluorouracil and metronidazole. **A.** *N*-nicotinoyl-2-acetoxy-D, L-glycine, a chemical drug delivery system for colonic delivery of drugs [50]. **B.** Proposed pathway for the bioactivation of *N*-nicotinoyl-2-(5-fluorouracil-1-yl)-D,L-glycine, a colon-specific prodrug of 5-fluorouracil [50]. **C.** *N*-nicotinoylglycyl-2-(5-fluorouracil-1-yl)-D,L-glycine, a colon-specific prodrug of 5-fluorouracil with another glycine between nicotinic acid and glycine of *N*-nicotinoyl-2-(5-fluorouracil-1-yl)-D, L-glycine [91]. **D.** *N*-Nicotinoyl-2-[2-(2-methyl-5-nitroimidazole-1-yl)ethoxy]glycine, a colon specific prodrug of metronidazole [44].

conversion rate becomes low. On the other hand, glucocorticoid sulfate ester conjugates were deconjugated at varying rates depending on the parent drugs. The order was prednisolone sulfate (PDS) > methylprednisolone sulfate (MPS) > dexamethasone sulfate (DS) in the cecal contents [81]. An *in vivo* study carried out with MPS and DS demonstrated no fecal recovery of the steroid prodrugs after oral administration [82,83]. However, it is not certain whether the difference in the extent of the prodrug conversion originates from the difference in conversion rates or the size of the dose, considering that oral doses of 5-ASA prodrugs are ~ 5 – 12 times those of steroid prodrugs under the assumption that a much longer time should be taken for complete conversion at larger doses [68,82,83].

For prodrugs with an azo bond, the electron density of the azo region affects the rate of the reductive bond cleavage.

A low electron density within the azo region facilitates reduction to form amine via the hydrazo intermediate. Thus, introduction of an electron-withdrawing group into an aromatic ring linked to an azo bond accelerates the azo bond cleavage [93].

The drug release from polymeric colon-specific prodrugs such as dextran-drug conjugates is initiated by the depolymerization of the polymer backbone, which determines the rate of drug release [73]. The dextranase-mediated depolymerization of dextran-5-ASA ester conjugates was attenuated compared with that of unsubstituted dextran, which depends on the degree of substitution and hydrophobicity of a drug coupled to dextran [71]. Accordingly, the candidate drug for the polymeric prodrug should either be hydrophilic or have high efficacy in order to provide effective dose on administration of a practical dose of prodrug. In other words, for the drug

with high efficacy (small dose), an effective dose might be achieved successively from a polymer drug with low degree of substitution. If the drug has low efficacy (large dose) the degree of substitution of the polymer drug should be high so that the effective dose will be provided from the polymer drug. In this case, the drug should be hydrophilic enough for facile bioconversion of the polymer drug.

In an *in vitro* experiment carried out with dextran-drug ester conjugates (degree of substitution: 20), treatment with dextranase for 8 h depolymerized the dextran-5-ASA ester conjugate up to 70%, whereas the dextran ester conjugates of flufenamic acid and 4-(4-ethoxycarbonylphenylazo)salicylic acid, which are more lipophilic than 5-ASA, were depolymerized up to 11 and 7.4%, respectively (unpublished data). Utilization of dextran as a colon-specific carrier seems to be suitable for hydrophilic drugs or for drugs whose effective doses are not high if they are hydrophobic.

In polymeric prodrugs with chemically and enzymatically stable backbones holding an azo bond in the pendant group as a colon-specific linkage, it seems that azo bonds are still cleavable because the reductive cleavage possibly takes place outside the bacterial cells by the small molecular reducing agents such as FMN and NADP, which are accessible to the azo bonds without the backbone degradation, functioning as a shuttle for the electron transport [31,94].

2.6.2 Metabolic stability of a parent drug in the colon

Metabolic stability of not only a colon-specific prodrug but also a parent drug could influence the regional therapeutic availability in the large intestine. The level of the parent drug at the target site should result from the net accumulation resulting from relative metabolic stability of a parent drug and its prodrug. The level of the drug depends mainly on the conversion rate of the prodrug if the parent drug is metabolically stable. In the other case, the drug should be at a certain level, which is determined by the difference between the rates of the prodrug conversion and the parent drug metabolism in the large intestine. Glucocorticoids are subject to reductive metabolism in the large intestine, which inactivates the drugs [95]. Kong *et al.* compared the relative metabolic stability of glucocorticoids and their sulfate prodrugs in the rat cecal contents and observed different accumulation profiles of drugs released from the prodrugs [81]. The accumulation of methylprednisolone and prednisolone, which are vulnerable to reductive metabolism, was lower than that of dexamethasone, which is resistant to it, in the rat cecal contents. The accumulation profiles (as a function of time) represented the net amounts of the steroids produced from corresponding prodrugs and disappeared metabolically. Consistent with this observation, in the large intestine the maximal concentration of dexamethasone after oral administration of DS (dose: 1 mg/kg) to rats was ~ 43 µg/ml, whereas that of methylprednisolone after MPS administration (dose: 2.36 mg/kg) was 45 µg/ml and the total amounts of dexamethasone and methylprednisolone accumulated for 8 h in the large intestine

were 630 and 250 µg, respectively [82,83]. Vulnerability of various glucocorticoids to reductive metabolism in the rat cecal contents was investigated [81,96]. Friend and Chang reported that the dexamethasone recovery in the cecum is much greater than that of prednisolone 4 – 5 h after oral administration of glucosides of prednisolone and dexamethasone, which is because of the difference in delivery efficiency of the two prodrugs [27]. However, the different susceptibility of glucocorticoids to the reductive metabolism suggests that the lower recovery of prednisolone comes from metabolic instability rather than delivery efficiency. A similar phenomenon was observed when rutin, a glycoside of quercetin, was incubated in the rat cecal contents. The natural product was deconjugated to liberate quercetin, which was metabolized by bacterial enzymes. The accumulation level of quercetin was much lower than for quercetin produced from rutin [51].

Metronidazole, an anti-amebic agent, is susceptible to reductive metabolism, which is relevant to bioactivation. Incubation of MTZ alone with the cecal contents resulted in rapid disappearance, indicating extensive metabolism of MTZ, which explains why MTZ was detected at a very low level compared with the disappearance of *N*-nicotinoyl-2-[2-(2-methyl-5-nitroimidazole-1-yl)ethoxy]glycine, a colon-specific MTZ prodrug, in the cecal contents on *in vitro* incubation or oral administration [44]. The authors suggest that a large fraction of MTZ produced from the prodrug was reduced to the active metabolite in the colonic bacteria, thus the observation that MTZ remained at a low level in the large intestine presents evidence that the prodrug should elicit effective antimicrobial activity.

Basit *et al.* reported that histamine receptor 2 blockers, a candidate drug for modification to a colon-specific prodrug, are metabolized rapidly by colonic bacteria. Ranitidine and nazatidine but not cimetidine and famotidine are susceptible to the colonic metabolism [97]. Other than the aforementioned drugs, 5-ASA, nalidixic acid, naproxen and ketoprofen do not seem to be metabolized significantly in the cecal contents. *N*-acetylation of 5-ASA was not detected on incubation with the cecal contents, whereas it was detected at a significant level in the colonic contents and the blood after oral administration of colon-specific 5-ASA prodrugs or 5-ASA [26,98,99], strongly suggesting 5-ASA metabolism in the colonic tissue and the liver. As it should affect therapeutic property of their colon-specific prodrugs, colonic metabolic stability of parent drugs needs to be taken into account for selecting a candidate drug on modification to a colon-specific prodrug.

3. Conclusion

This article has reviewed what should be considered on designing a colon-specific drug delivery system using prodrug approaches. As mentioned, the requirement for colonic delivery of a drug can be satisfied by various chemical modifications of the drug, which make it hydrophilic or large enough to prevent systemic absorption in the upper intestine, and can be

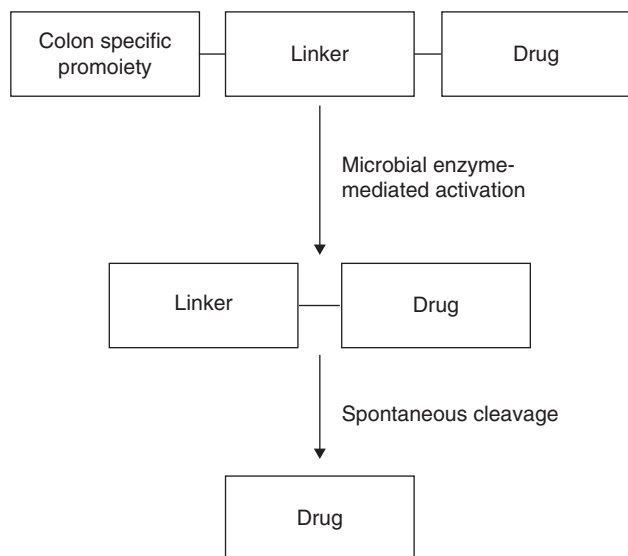


Figure 4. General principle of the bioactivation of a colon-specific chemical drug delivery system.

restored to the intact drug by microbial enzymes in the large intestine but not by host ones in the upper intestine. Prodrug approaches are a very efficient way to deliver a drug to the last part of the GI tract, which provides therapeutic advantages for treatment of colonic diseases, protein and peptide therapy and chronotherapy. However, many obstacles should be overcome to develop a clinically useful colon-specific prodrug. Chemical modification of a drug entails safety issues and requires an appropriate functional group that may not exist in all candidate drugs. In addition, a difference in colonic microflora among individuals and even in a single subject is likely to result in variation in therapeutic effects of colon-specific prodrugs. Colonic distribution of the parent drug after oral administration of a colon-specific prodrug is relevant to an efficient therapeutic action against a colonic disease whose pathogenic lesion occurs not only in the proximal part but also in the distal part of the large intestine. The regional therapeutic availability at the target site depends on colonic metabolic stability of the prodrug and the parent drug. On design of a colon-specific prodrug, the factors that influence therapeutic activity and reproducibility should be taken into consideration, which could be controlled by various tactics that have been developed and extensive knowledge that has been accumulated in the field of colon-specific drug delivery using prodrug approaches.

4. Expert opinion

The term prodrug was first introduced by Albert in 1958 to describe compounds that must undergo a chemical transformation within the body to elicit therapeutic action. However, it was not until the early 1970s that worldwide attention was directed to this area as a potential chemical approach to

improving drug therapy [100]. Over the past 30 years since then, this technique has undergone considerable expansion, largely as a result of the increased awareness and understanding of the physicochemical and biological factors that determine the efficacy of prodrugs, which has led the role of prodrugs to increase in modern drug discovery programs. Of the total of 49 new chemical entities approved in the United States in 2001 and 2002, prodrugs occupied 22 – 24% of the new drugs. A recent analysis of a listing of all drugs approved worldwide estimated that 5 – 7% were in fact prodrugs. Considering that the number of prodrug patents and papers searched in PubMed and SciFinder each year has increased significantly over the last 10 years (1993 – 2003), it is likely that the portion of prodrugs in the new chemical entities approved will grow in the future [101].

Prodrug approaches are a feasible way to deliver a drug specifically to the large intestine, probably leading to improved pharmacological and toxicological properties of the drug. The great specificity in delivery and the potential for patent protection can act as a driving force to developing a colon-specific prodrug. On the other hand, there are several barriers to be overcome for the development of clinically useful colon-specific prodrugs. Entailing chemical modification of a drug, prodrugs are considered to be new molecular entities, which raises issues on safety. Before reaching the market, the prodrugs must come through animal and human studies on safety [20]. The requirement for functional groups in drugs for chemical modification puts a limitation on the general application of the approach to drugs. Other than the seem-to-be-inborn disadvantages, inter- and intra-individual variation in prodrug activation and the colonic metabolism of parent drugs may add difficulty in developing a practical colon-specific prodrug. One strategy to resolve such obstacles is to have various colon-specific promoieties coupling with a functional group. Drugs for which hydroxyl group is available could be modified to glycosides, sulfate esters, dextran-drug ester conjugates and *N*-nicotinoyl-(drug-oxy)-DL-glycine [25,27,44,84]. Conversion of the colon-specific prodrugs to the drugs is carried out by different activation systems (bacterial enzymes) in the large intestine, which may provide an option minimizing variation in the prodrug activation and controlling the colonic distribution of the active drugs. Development of a new colon-specific promoiety should make a contribution not only to running over the hurdles in achieving a reproducible clinical effect, but also to lessening the limitation of the prodrug approach set by the requirement of a functional group. The functional group limitation can be categorized into two cases. The first category includes drugs with no available functional group. The second one includes drugs with functional groups able to couple with a colon-specific promoiety, where the bonds are not cleaved (effectively) by colonic bacterial enzymes owing to steric hindrance and/or no susceptibility to the enzymes. For the drugs in the first category, pharmaceutical formulation would be the first option for colonic delivery [21]. A smart strategy to circumvent

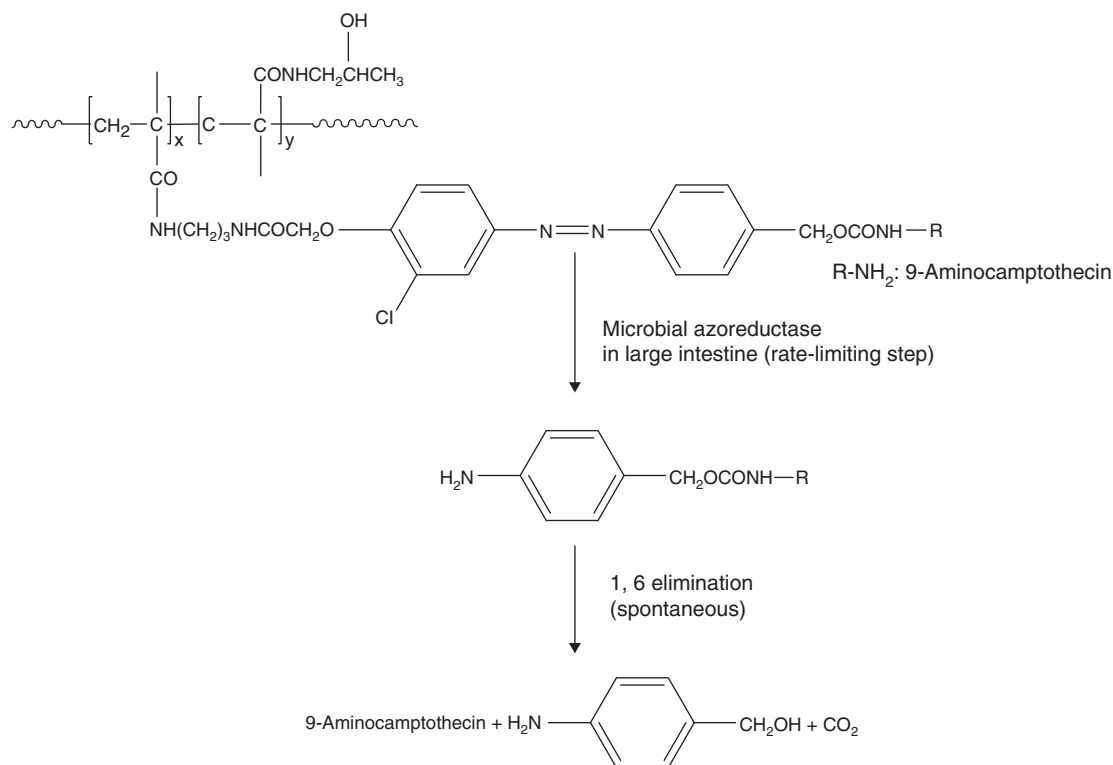


Figure 5. The bioactivation pathway of a polymeric colon specific-prodrug of 9-aminocamptothecin with azo bond and a self-1,6 eliminating linker in the pendant group [102].

the drawback derived from the second is the introduction of a chemical drug delivery system. The chemical drug delivery system generally refers to the use of pro-prodrugs, and the general principle of a colon-specific chemical drug delivery system is illustrated in Figure 4. The colon-specific cleavage takes place between the colon-specific promoiety and the linker to form intermediate drug-linker derivative that decomposes spontaneously to liberate the parent drug. The linker acts as a spacer to avoid steric hindrance between the linker and the promoiety and the spontaneous liberation of the parent drugs from the linker acts as a mechanism to circumvent no (efficient) enzymatic liberation of the parent drug. Lee *et al.* adopted the pro-prodrug concept to colon-specific prodrug (of 5-FU) for the first time, as shown in Figure 3B [50]. Gao *et al.* used a more elaborate chemical delivery system with a polymeric colon-specific prodrug, where a self-eliminating linker functioning with 1,6 elimination chemistry was used as a spacer between HPMA copolymer and 9-aminocamptothecin (9-AC) and was triggered by colon-specific azo reduction, resulting in spontaneous liberation of 9-AC, as shown in Figure 5 [102]. This polymeric prodrug demonstrated enhanced 9-AC release compared with a polymeric prodrug of 9-AC using an aromatic azo bond and dipeptide as a spacer that requires the enzymatic cleavage between 9-AC and an amino acid residue to liberate 9-AC

following the reduction of the azo bond [32]. Regarding colonic metabolism of parent drugs that could result in therapeutic variation and loss of their colon-specific prodrugs, replacing such a drug with a metabolically stable one would be the first choice to resolve the problem. For example, prednisolone, which is susceptible to reductive metabolism (bioinactivation), could be substituted with metabolically stable dexamethasone on selection of a glucocorticoid as a parent drug for modification to a colon-specific prodrug. As an alternative, it could be considered to co-administer enzyme inhibitors. The possibility of this method was shown in a paper demonstrating that reduction of metronidazole was partly prevented in the presence of reduction inhibitors in the cecal contents [44]. To avoid side effects and loss in the upper intestine on co-administration of enzyme inhibitors, the inhibitors should also be colon-specific. In this sense, natural glycosides such as rutin and glycyrrhizin, which possess both colon specificity and inhibitory activity on reductase [51,103,104], could be applicable to co-administration with reductase-sensitive glucocorticoids including prednisolone for treatment of IBD. There may be advantages of co-administration over replacement with a reductase-resistant glucocorticoid (dexamethasone). As prednisolone is much more vulnerable to metabolism in the liver than dexamethasone [81], co-administration, which may stabilize prednisolone at the colon but not in the liver,

could increase therapeutic concentration at the target site but not in the blood, probably resulting in a lower possibility of systemic side effects than replacement with dexamethasone. Another advantage is that the glycoside reductase inhibitors, which have an anti-inflammatory effect on experimental colitis [51,105], may potentiate the anti-inflammatory effect of glucocorticoids against IBD on co-administration. Although delivering drugs to the last part of the GI tract, the large intestine, is not an easy task, colonic drug delivery is definitely worth all the trouble considering its therapeutic advantages. Prodrug approaches, an attractive option for colonic drug delivery, can be extended by diversifying and organizing tactics to provide colon-targetability for drugs, wherein the strategy based on

chemistry would make a greater contribution to the site-specific drug delivery.

Acknowledgements

Both authors contributed equally to the preparation of this review.

Declaration of interest

This work was supported by the Korea Research Foundation Grants funded by the Korean Government (KRF-2007-314-E00245) and (KRF-2006-531-E00116).

Bibliography

Papers of special note have been highlighted as either of interest (•) or of considerable interest (••) to readers.

- Braun K, Pipkorn R, Waldeck W. Development and characterization of drug delivery systems for targeting mammalian cells and tissues: a review. *Curr Med Chem* 2005;12:1841-58
- Langer R. Drug delivery and targeting. *Nature* 1998;392:5-10
- Ruoslahti E. Drug targeting to specific vascular sites. *Drug Discov Today* 2002;7:1138-43
- Ast G. Drug-targeting strategies for prostate cancer. *Curr Pharm Des* 2003;9:455-66
- Haas M, Moolenaar F, Meijer DK, et al. Specific drug delivery to the kidney. *Cardiovasc Drugs Ther* 2002;16:489-96
- de Boer AG, Gaillard PJ. Drug targeting to the brain. *Annu Rev Pharmacol Toxicol* 2007;47:323-55
- Takahashi-Nishioka T, Yokogawa K, Tomatsu S, et al. Targeted drug delivery to bone: pharmacokinetic and pharmacological properties of acidic oligopeptide-tagged drugs. *Curr Drug Discov Technol* 2008;5:39-48
- Wilding I. Site-specific drug delivery in the gastrointestinal tract. *Crit Rev Ther Drug Carrier Syst* 2000;17:557-620
- Pavan B, Dalpiaz A, Ciliberti N, et al. Progress in drug delivery to the central nervous system by the prodrug approach. *Molecules* 2008;13:1035-65
- Shareef MA, Khar RK, Ahuja A, et al. Colonic drug delivery: an updated review. *AAPS PharmSci* 2003;5:E17
- Singh Y, Palombo M, Sinko PJ. Recent trends in targeted anticancer prodrug and conjugate design. *Curr Med Chem* 2008;15:1802-26
- Jain SK, Jain A. Target-specific drug release to the colon. *Expert Opin Drug Deliv* 2008;5:483-98
- Patel M, Shah T, Amin A. Therapeutic opportunities in colon-specific drug-delivery systems. *Crit Rev Ther Drug Carrier Syst* 2007;24:147-202
- Friend DR. Review article: issues in oral administration of locally acting glucocorticosteroids for treatment of inflammatory bowel disease. *Aliment Pharmacol Ther* 1998;12:591-603
- Sinha V, Singh A, Kumar RV, et al. Oral colon-specific drug delivery of protein and peptide drugs. *Crit Rev Ther Drug Carrier Syst* 2007;24:63-92
- Saffran M, Kumar GS, Savariar C, et al. A new approach to the oral administration of insulin and other peptide drugs. *Science* 1986;233:1081-4
- **The first paper on colonic delivery of protein and peptide drugs using an azo polymer.**
- Bussemer T, Otto I, Bodmeier R. Pulsatile drug-delivery systems. *Crit Rev Ther Drug Carrier Syst* 2001;18:433-58
- Kinget R, Kalala W, Vervoort L, et al. Colonic drug targeting. *J Drug Target* 1998;6:129-49
- Van den Mooter G. Colon drug delivery. *Expert Opin Drug Deliv* 2006;3:111-25
- Friend DR. New oral delivery systems for treatment of inflammatory bowel disease. *Adv Drug Deliv Rev* 2005;57:247-65
- **A well-organized review on colon-specific delivery of drugs for treatment of inflammatory bowel disease.**
- Chourasia MK, Jain SK. Pharmaceutical approaches to colon targeted drug delivery systems. *J Pharm Pharm Sci* 2003;6:33-66
- Sinha VR, Kumria R. Colonic drug delivery: prodrug approach. *Pharm Res* 2001;18:557-64
- **An excellent review on colon-specific prodrugs.**
- Chae SY, Jang MK, Nah JW. Influence of molecular weight on oral absorption of water soluble chitosans. *J Control Release* 2005;102:383-94
- Ungell AL, Nylander S, Bergstrand S, et al. Membrane transport of drugs in different regions of the intestinal tract of the rat. *J Pharm Sci* 1998;87:360-6
- Doh MJ, Jung YJ, Kim I, et al. Synthesis and in vitro properties of prednisolone 21-sulfate sodium as a colon-specific prodrug of prednisolone. *Arch Pharm Res* 2003;26:258-63
- Jung YJ, Lee JS, Kim YM. Synthesis and in vitro/in vivo evaluation of 5-aminosalicyl-glycine as a colon-specific prodrug of 5-aminosalicylic acid. *J Pharm Sci* 2000;89:594-602
- **Paper on the *in vitro* and *in vivo* properties of a colon-specific prodrug of 5-aminosalicylic acid whose amine is conjugated with glycine by means of an amide bond.**
- Friend DR, Chang GW. A colon-specific drug-delivery system based on drug

What should be considered on design of a colon-specific prodrug?

- glycosides and the glycosidases of colonic bacteria. *J Med Chem* 1984;27:261-6
- **Synthesis and colon specificity of glucose-conjugated prednisolone and dexamethasone (steroid glycosides).**
- 28. Minami K, Hirayama F, Uekama K. Colon-specific drug delivery based on a cyclodextrin prodrug: release behavior of biphenylacetic acid from its cyclodextrin conjugates in rat intestinal tracts after oral administration. *J Pharm Sci* 1998;87:715-20
- 29. Harboe E, Larsen C, Johansen M, et al. Macromolecular prodrugs. XV. Colon-targeted delivery-bioavailability of naproxen from orally administered dextran-naproxen ester prodrugs varying in molecular size in the pig. *Pharm Res* 1989;6:919-23
- **Dextran-naproxen ester conjugates were evaluated as a colon-specific prodrug for colonic absorption of naproxen by investigating pharmacokinetics after oral administration of the prodrugs varying in molecular size.**
- 30. Leopold CS, Friend DR. In vivo pharmacokinetic study for the assessment of poly(L-aspartic acid) as a drug carrier for colon-specific drug delivery. *J Pharmacokinet Biopharm* 1995;23:397-406
- 31. Brown JP, McGarraugh GV, Parkinson TM, et al. A polymeric drug for treatment of inflammatory bowel disease. *J Med Chem* 1983;26:1300-7
- **A non-biodegradable polymer with 5-aminosalicylic acid coupled to its pendant group via an azo bond was evaluated as a colon-specific prodrug of 5-aminosalicylic acid.**
- 32. Sakuma S, Lu ZR, Kopeckova P, et al. Biorecognizable HPMA copolymer-drug conjugates for colon-specific delivery of 9-aminocamptothecin. *J Control Release* 2001;75:365-79
- 33. Wiwattanapatapee R, Lomlim L, Saramunee K. Dendrimers conjugates for colonic delivery of 5-aminosalicylic acid. *J Control Release* 2003;88:1-9
- 34. Scheline RR. Metabolism of foreign compounds by gastrointestinal microorganisms. *Pharmacol Rev* 1973;25:451-523
- **An old review on metabolism of foreign compounds by intestinal microflora, which is still very informative for the design of a colon-specific prodrug.**
- 35. Haeberlin B. FDR. Anatomy and physiology of the gastrointestinal tract: Implication for colonic drug delivery. In: Friend DR, editor. *Oral colon-specific drug delivery*. CRC Press, Inc., Boca Raton, FL; 1992. p. 1-43
- **An excellent book including almost all of the information for oral colon-specific drug delivery.**
- 36. Brown JP. Reduction of polymeric azo and nitro dyes by intestinal bacteria. *Appl Environ Microbiol* 1981;41:1283-6
- 37. Faigle JW. Drug metabolism in the colon wall and lumen. In: Bieck PR, editor. *Colonic drug absorption and metabolism*. Marcel Dekker, Inc.; 1993. p. 29-54
- 38. Sinha VR, Kumria R. Microbially triggered drug delivery to the colon. *Eur J Pharm Sci* 2003;18:3-18
- 39. Guarner F, Malagelada JR. Gut flora in health and disease. *Lancet* 2003;361:512-9
- 40. Keighley MR, Arabi Y, Dimock F, et al. Influence of inflammatory bowel disease on intestinal microflora. *Gut* 1978;19:1099-104
- 41. Carrette O, Favier C, Mizon C, et al. Bacterial enzymes used for colon-specific drug delivery are decreased in active Crohn's disease. *Dig Dis Sci* 1995;40:2641-6
- 42. Bhat P, Shantakumari S, Rajan D, et al. Bacterial flora of the gastrointestinal tract in southern Indian control subjects and patients with tropical sprue. *Gastroenterology* 1972;62:11-21
- 43. Egan LJ, Sandborn WJ. Drug therapy of inflammatory bowel disease. *Drugs Today (Barc)* 1998;34:431-46
- 44. Kim D, Hong S, Jung S, et al. Synthesis and Evaluation of N-Nicotinoyl-2-[2-(2-Methyl-5-Nitroimidazol-1-yl) Ethyloxy]-D,L-Glycine as a Colon-Specific Prodrug of Metronidazole. *J Pharm Sci* 2009;98(11):4161-9
- 45. Lee JS, Jung YJ, Doh MJ, et al. Synthesis and properties of dextran-nalidixic acid ester as a colon-specific prodrug of nalidixic acid. *Drug Dev Ind Pharm* 2001;27:331-6
- 46. Pradhan A, Majumdar MK. Metabolism of some drugs by intestinal lactobacilli and their toxicological considerations. *Acta Pharmacol Toxicol (Copenh)* 1986;58:11-5
- 47. Nolen HW III, Friend DR. Menthol-beta-D-glucuronide: a potential prodrug for treatment of the irritable bowel syndrome. *Pharm Res* 1994;11:1707-11
- 48. Simpkins JW, Smulkowski M, Dixon R, et al. Evidence for the delivery of narcotic antagonists to the colon as their glucuronide conjugates. *J Pharmacol Exp Ther* 1988;244:195-205
- 49. Rodrigues CM, Kren BT, Steer CJ, et al. The site-specific delivery of ursodeoxycholic acid to the rat colon by sulfate conjugation. *Gastroenterology* 1995;109:1835-44
- **O-sulfation was evaluated as a colon-specific linkage and the therapeutic advantage of colonic delivery of ursodeoxycholic acid was investigated.**
- 50. Lee JS, Jung YJ, Kim YM. Synthesis and evaluation of N-acyl-2-(5-fluorouracil-1-yl)-D,L-glycine as a colon-specific prodrug of 5-fluorouracil. *J Pharm Sci* 2001;90:1787-94
- **A chemical drug delivery system (pro-prodrug) was applied to a colon-specific delivery of 5-fluorouracil.**
- 51. Kim H, Kong H, Choi B, et al. Metabolic and pharmacological properties of rutin, a dietary quercetin glycoside, for treatment of inflammatory bowel disease. *Pharm Res* 2005;22:1499-509
- 52. Kobashi K, Nishimura T, Kusaka M, et al. Metabolism of sennosides by human intestinal bacteria. *Planta Med* 1980;40:225-36
- 53. Krishnaiah YS, Veer Raju P, Dinesh Kumar B, et al. Development of colon targeted drug delivery systems for mebendazole. *J Control Release* 2001;77:87-95
- 54. Krishnaiah YS, Satyanarayana V, Kumar BD, et al. Studies on the development of colon-targeted delivery systems for celecoxib in the prevention of colorectal cancer. *J Drug Target* 2002;10:247-54
- 55. Sinha VR, Bhinge JR, Kumria R, et al. Development of pulsatile systems for targeted drug delivery of celecoxib for prophylaxis of colorectal cancer. *Drug Deliv* 2006;13:221-5
- 56. Tozaki H, Nishioka J, Komoike J, et al. Enhanced absorption of insulin and (Asu(1,7))eel-calcitonin using novel azopolymer-coated pellets for colon-specific drug delivery. *J Pharm Sci* 2001;90:89-97

57. Tozaki H, Komoike J, Tada C, et al. Chitosan capsules for colon-specific drug delivery: improvement of insulin absorption from the rat colon. *J Pharm Sci* 1997;86:1016-21
58. Van den Mooter G, Maris B, Samyn C, et al. Use of azo polymers for colon-specific drug delivery. *J Pharm Sci* 1997;86:1321-7
59. Larsen C, Jensen BH, Olesen HP. Bioavailability of ketoprofen from orally administered ketoprofen-dextran ester prodrugs in the pig. *Acta Pharm Nord* 1991;3:71-6
60. Robinson MG. New oral salicylates in the therapy of chronic idiopathic inflammatory bowel disease. *Gastroenterol Clin North Am* 1989;18:43-50
61. Chan RP, Pope DJ, Gilbert AP, et al. Studies of two novel sulfasalazine analogs, ipsalazide and balsalazide. *Dig Dis Sci* 1983;28:609-15
62. Yamaguchi T, Sasaki K, Kurosaki Y, et al. Biopharmaceutical evaluation of salicylazosulfanilic acid as a novel colon-targeted prodrug of 5-aminosalicylic acid. *J Drug Target* 1994;2:123-31
63. Dhaneshwar SS, Gairola N, Kandpal M, et al. Synthesis, kinetic studies and pharmacological evaluation of mutual azo prodrug of 5-aminosalicylic acid with D-phenylalanine for colon specific drug delivery in inflammatory bowel disease. *Bioorg Med Chem Lett* 2007;17:1897-902
- **The 5-aminosalicylic acid derivative where the phenylalanine is linked to the aromatic amine via an azo bond was evaluated as a mutual colon-specific prodrug. Phenylalanine acted as not only a colon-specific carrier but also a therapeutic agent enhancing the anti-inflammatory effect of 5-aminosalicylic acid.**
64. Lu ZR, Shiah JG, Sakuma S, et al. Design of novel bioconjugates for targeted drug delivery. *J Control Release* 2002;78:165-73
65. Canevari M, Castagliuolo I, Brun P, et al. Poly(ethylene glycol)-mesalazine conjugate for colon specific delivery. *Int J Pharm* 2009;368:171-7
66. Davaran S, Hanaee J, Khosravi A. Release of 5-amino salicylic acid from acrylic type polymeric prodrugs designed for colon-specific drug delivery. *J Control Release* 1999;58:279-87
67. Pellicciari R, Garzon-Aburbeh A, Natalini B, et al. Brush-border-enzyme-mediated intestine-specific drug delivery. Amino acid prodrugs of 5-aminosalicylic acid. *J Med Chem* 1993;36:4201-7
68. Jung Y, Kim HH, Kim H, et al. Evaluation of 5-aminosalicyltaurine as a colon-specific prodrug of 5-aminosalicylic acid for treatment of experimental colitis. *Eur J Pharm Sci* 2006;28:26-33
- **The 5-aminosalicylic acid derivative where the taurine is linked to the carboxylic acid of 5-ASA via an amide bond was evaluated as a dual colon-specific prodrug. Taurine acted as not only a colon-specific carrier but also a therapeutic agent potentiating the anti-inflammatory effect of 5-aminosalicylic acid.**
69. Kim H, Huh J, Jeon H, et al. N,N'-Bis (5-aminosalicyl)-L-cystine is a potential colon-specific 5-aminosalicylic acid prodrug with dual therapeutic effects in experimental colitis. *J Pharm Sci* 2009;98:159-68
70. Kim H, Jeon H, Kong H, et al. A molecular mechanism for the anti-inflammatory effect of taurine-conjugated 5-aminosalicylic acid in inflamed colon. *Mol Pharmacol* 2006;69:1405-12
- **This paper investigated a molecular mechanism underlying taurine potentiation of the anti-inflammatory effect of 5-ASA against experimental colitis.**
71. Jung YJ, Lee JS, Kim HH, et al. Synthesis and properties of dextran-5-aminosalicylic acid ester as a potential colon-specific prodrug of 5-aminosalicylic acid. *Arch Pharm Res* 1998;21:179-86
72. Inoue M, Morikawa M, Tsuboi M, et al. Species difference and characterization of intestinal esterase on the hydrolyzing activity of ester-type drugs. *Jpn J Pharmacol* 1979;29:9-16
73. Larsen C, Harboe E, Johansen M, et al. Macromolecular prodrugs. XVI. Colon-targeted delivery—comparison of the rate of release of naproxen from dextran ester prodrugs in homogenates of various segments of the pig gastrointestinal (GI) tract. *Pharm Res* 1989;6:995-9
- **This paper provides a rationale on colon specificity of dextran-drug ester prodrugs.**
74. Herzog R, Leuschner J. Experimental studies on the pharmacokinetics and toxicity of 5-aminosalicylic acid-O-sulfate following local and systemic application. *Arzneimittelforschung* 1995;45:300-3
75. Crotty B, Jewell DP. Drug therapy of ulcerative colitis. *Br J Clin Pharmacol* 1992;34:189-98
76. Friend DR, Chang GW. Drug glycosides: potential prodrugs for colon-specific drug delivery. *J Med Chem* 1985;28:51-7
- **The influence of prodrug structure on specificity of glycoside/glycosidase-based colon-specific drug delivery was studied by preparing nine steroid glycosides.**
77. Conchie J, Macdonald DC. Glycosidases in the mammalian alimentary tract. *Nature* 1959;184(Suppl 16):1233
78. Haeblerlin B, Rubas W, Nolen HW III, et al. In vitro evaluation of dexamethasone-beta-D-glucuronide for colon-specific drug delivery. *Pharm Res* 1993;10:1553-62
- **Dexamethasone-b-D-glucuronide was *in vitro*-evaluated as a potential colon-specific prodrug of the glucocorticoids for treatment of inflammatory bowel disease.**
79. Cui N, Friend DR, Fedorak RN. A budesonide prodrug accelerates treatment of colitis in rats. *Gut* 1994;35:1439-46
80. Jung YJ, Doh MJ, Kim IH, et al. Prednisolone 21-sulfate sodium: a colon-specific pro-drug of prednisolone. *J Pharm Pharmacol* 2003;55:1075-82
- **An *in vitro* and *in vivo* evaluation of prednisolone 21-sulfate sodium as a colon-specific prodrug of prednisolone.**
81. Kong H, Kim Y, Lee Y, et al. Sulfate-conjugated methylprednisolone: evaluation as a colon-specific methylprednisolone prodrug and comparison with sulfate-conjugated prednisolone and dexamethasone. *J Drug Target* 2009;17:159-67
- **This paper shows that the accumulation profiles of the glucocorticoids on incubation of three glucocorticoid prodrugs with the cecal contents depends on the metabolic stability of the free glucocorticoids.**
82. Kim I, Kong H, Lee Y, et al. Dexamethasone 21-sulfate improves the therapeutic properties of dexamethasone against experimental rat colitis by

What should be considered on design of a colon-specific prodrug?

- specifically delivering the steroid to the large intestine. *Pharm Res* 2009;26:415-21
- **A colon-specific prodrug of dexamethasone, dexamethasone 21-sulfate, increased potency and reduced adverse effects of dexamethasone on oral administration for treatment of rat colitis.**
83. Kong H, Lee Y, Hong S, et al. Sulfate-conjugated methylprednisolone as a colon-targeted methylprednisolone prodrug with improved therapeutic properties against rat colitis. *J Drug Target* 2009;17:450-8
 84. McLeod AD, Friend DR, Tozer TN. Glucocorticoid-dextran conjugates as potential prodrugs for colon-specific delivery: hydrolysis in rat gastrointestinal tract contents. *J Pharm Sci* 1994;83:1284-8
 - **An *in vitro* evaluation of glucocorticoid-succinic acid-dextran conjugates as a polymeric colon-specific prodrug of the glucocorticoids.**
 85. McLeod AD, Fedorak RN, Friend DR, et al. A glucocorticoid prodrug facilitates normal mucosal function in rat colitis without adrenal suppression. *Gastroenterology* 1994;106:405-13
 - **This paper shows that glucocorticoid-succinic acid-dextran conjugates can be administered orally to facilitate mucosal repair in rat colitis without adenosuppression.**
 86. Yano H, Hirayama F, Arima H, et al. Preparation of prednisolone-appended alpha-, beta- and gamma-cyclodextrins: substitution at secondary hydroxyl groups and *in vitro* hydrolysis behavior. *J Pharm Sci* 2001;90:493-503
 - **Cyclodextrins were tested as a colon-specific carrier of a glucocorticoid.**
 87. Yano H, Hirayama F, Kamada M, et al. Colon-specific delivery of prednisolone-appended alpha-cyclodextrin conjugate: alleviation of systemic side effect after oral administration. *J Control Release* 2002;79:103-12
 - **Therapeutic and toxicological advantages of prednisolone-appended α -cyclodextrin conjugate over prednisolone in treatment of rat colitis.**
 88. Diasio RB, Harris BE. Clinical pharmacology of 5-fluorouracil. *Clin Pharmacokinet* 1989;16:215-37
 89. Mukherjee KL, Heidelberger C. Studies on fluorinated pyrimidines. IX. The degradation of 5-fluorouracil-6-C14. *J Biol Chem* 1960;235:433-7
 90. Ho DH, Townsend L, Luna MA, et al. Distribution and inhibition of dihydrouracil dehydrogenase activities in human tissues using 5-fluorouracil as a substrate. *Anticancer Res* 1986;6:781-4
 91. Lee J, Rho J, Yang Y, et al. Synthesis and *in vitro* evaluation of N-nicotinoylglycyl-2-(5-fluorouracil-1-yl)-D,L-glycine as a colon-specific prodrug of 5-fluorouracil. *J Drug Target* 2007;15:199-205
 - **Another glycine was inserted between nicotinic acid and glycine of N-acyl-2-(5-fluorouracil-1-yl)-DL-glycine [49], which increases 5-fluorouracil release in the cecal contents.**
 92. Jung YJ, Kim HH, Kong HS, et al. Synthesis and properties of 5-aminosalicyl- α -taurine as a colon-specific prodrug of 5-aminosalicylic acid. *Arch Pharm Res* 2003;26:264-9
 93. Brondsted H, Kopecek J. Hydrogels for site-specific drug delivery to the colon: *in vitro* and *in vivo* degradation. *Pharm Res* 1992;9:1540-5
 94. Dubin P, Wright KL. Reduction of azo food dyes in cultures of *Proteus vulgaris*. *Xenobiotica* 1975;5:563-71
 95. Shamat MA. The role of the gastrointestinal microflora in the metabolism of drugs. *Int J Pharm* 1993;97:1-13
 96. Kim IH, Kong HS, Choi BI, et al. Synthesis and *in vitro* properties of dexamethasone 21-sulfate sodium as a colon-specific prodrug of dexamethasone. *Drug Dev Ind Pharm* 2006;32:389-97
 97. Basit AW, Newton JM, Lacey LF. Susceptibility of the H2-receptor antagonists cimetidine, famotidine and nizatidine, to metabolism by the gastrointestinal microflora. *Int J Pharm* 2002; 237:23-33
 98. Jung YJ, Lee JS, Kim YM. Colon-specific prodrugs of 5-aminosalicylic acid: synthesis and *in vitro/in vivo* properties of acidic amino acid derivatives of 5-aminosalicylic acid. *J Pharm Sci* 2001;90:1767-75
 99. Zhou SY, Fleisher D, Pao LH, et al. Intestinal metabolism and transport of 5-aminosalicylate. *Drug Metab Dispos* 1999;27:479-85
 - **An excellent paper on intestinal absorption and metabolism of 5-aminosalicylic acid.**
 100. Christrup LL, Moss J, Steffansen B. Prodrugs. In: Swarbrick J, Boylan JC, editors. *Encyclopedia of pharmaceutical technology*. Volume 13. Marcel Dekker, Inc.; 1988. p. 39-70
 101. Stella VJ. Case for prodrugs. In: Stella VJ, Borchardt RT, Hageman MJ, et al., editors. *Prodrugs: challenges and rewards*. New York: Springer Science; 2007. p. 3-36
 102. Gao SQ, Lu ZR, Petri B, et al. Colon-specific 9-aminocamptothecin-HPMA copolymer conjugates containing a 1,6-elimination spacer. *J Control Release* 2006;110:323-31
 - **An *in vitro* investigation on a polymeric colon-specific prodrug of 5-aminosalicylic acid utilizing spontaneous 1,6 elimination following azo reduction as a drug release mechanism.**
 103. Hiipakka RA, Zhang HZ, Dai W, et al. Structure-activity relationships for inhibition of human 5 α -reductases by polyphenols. *Biochem Pharmacol* 2002;63:1165-76
 104. Ojima M, Satoh K, Gomibuchi T, et al. The inhibitory effects of glycyrrhizin and glycyrrhetic acid on the metabolism of cortisol and prednisolone—*in vivo* and *in vitro* studies. *Nippon Naibunpi Gakkai Zasshi* 1990;66:584-96
 105. Yuan H, Ji WS, Wu KX, et al. Anti-inflammatory effect of Diammonium Glycyrrhizinate in a rat model of ulcerative colitis. *World J Gastroenterol* 2006;12:4578-81

Affiliation

Yunjin Jung[†] & Young Mi Kim
[†]Author for correspondence
 Pusan National University,
 College of Pharmacy,
 Laboratory of Biomedical/
 Medicinal Chemistry,
 Busan, 609-735 Korea
 Tel: +051 510 2527; Fax: +051 513 6754;
 E-mail: jungy@pusan.ac.kr